

Access to Six- and Seven-Membered 1,7-Fused Indolines via Rh(III)-Catalyzed Redox-Neutral C7-Selective C–H Functionalization of Indolines with Alkynes and Alkenes

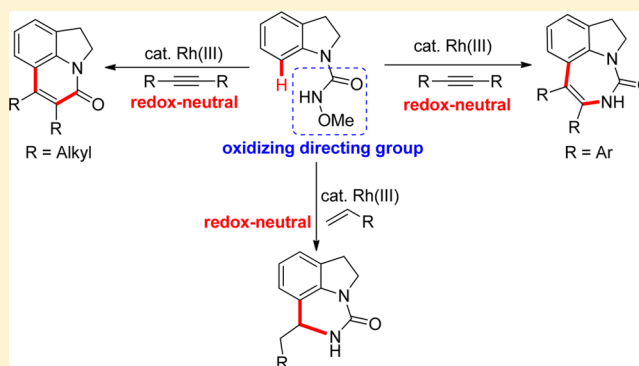
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S Supporting Information

ABSTRACT: We report herein a new strategy for the Rh(III)-catalyzed redox-neutral C7-selective C–H activation/annulation of indolines to rapidly access various privileged 1,7-fused indolines by utilizing an oxidizing–directing group. For example, a Rh(III)-catalyzed redox-neutral C7-selective C–H functionalization of indolines with arylalkynes is described to directly access 7-membered 1,7-fused indolines. Moreover, an unprecedented intramolecular addition of an alkenyl-Cp*Rh(III) species to a carbamoyl moiety occurred to give 1*H*-pyrroloquinolinones when employing alkyl alkynes. Additionally, an efficient Rh(III)-catalyzed redox-neutral C7-selective C–H activation/alkenylation/aza-Michael addition of indolines is also developed to give 6-membered 1,7-fused indolines.

The advantages of these processes are as follows: (1) mild and simple reaction conditions; (2) no need for an external oxidant; (3) broad scope of substrates; and (4) valuable six- or seven-membered 1,7-fused indolines as products.



INTRODUCTION

The indoles and indolines have been a topic of substantial research interest due to their ubiquity in numerous natural bioactive products, marketed drugs, pharmaceutically important compounds, and other functional molecules.¹ Among them, 1,7-fused indoles and indolines are particularly noteworthy due to their prevalence in numerous natural bioactive products and pharmaceutically important compounds (Figure 1).² However, 1,7-fused indolines are typically formed via an intramolecular

fashion. Moreover, the annulation precursors are normally not readily available and require multistep synthesis.³ To date, only one intermolecular catalytic method is available to directly access 1,7-fused indoline core. However, this method requires the installation of 2 equiv of terminal alkynes, thus largely limiting the diversity of the products.⁴ Therefore, the development of general synthetic methods for rapid preparation of 1,7-fused indolines and indoles would be highly desirable.

On the other hand, with the development of catalytic C–H functionalization, direct C–H bond functionalization of indoles and indolines should be one of the most effective and straightforward approaches to access substituted indoles and indolines.⁵ Although several elegant approaches to transition-metal-catalyzed C2- or C3-H functionalization of indoles have been reported,^{6,7} the indole C7 C–H functionalization has very limited reports.⁸ Recently, some examples of transition-metal-catalyzed chelation-assisted direct C7 C–H functionalization of indolines, such as arylation,⁹ alkenylation,¹⁰ alkylation,¹¹ amidation,¹² acylation,¹³ and alkylation,¹⁴ have been disclosed by several groups and us (Scheme 1a). However, no 1,7-fused indolines could be accessed through these methods.

Recently, the Rh(III)-catalyzed redox-neutral functionalization of aryl C–H bond with alkynes and alkenes has proven to be a powerful method for the rapid assembly of various complex molecular structures under mild reaction conditions without the use of external oxidant by utilizing an oxidizing–directing

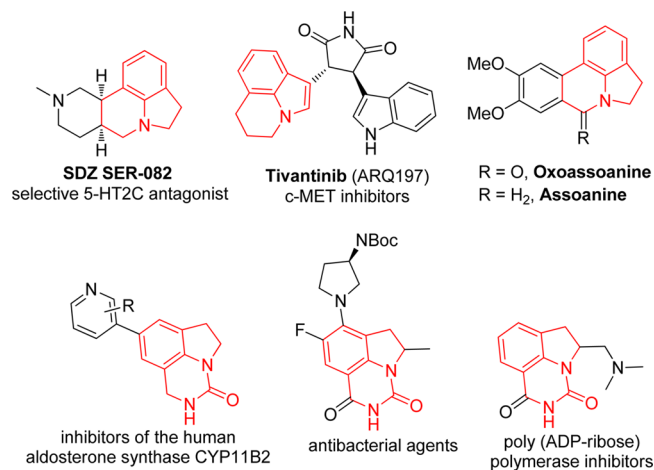


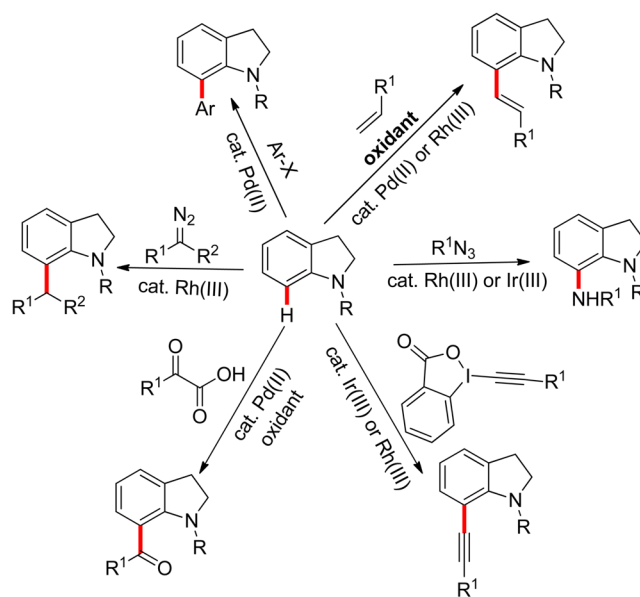
Figure 1. Selective bioactive compounds based on 1,7-fused indolines.

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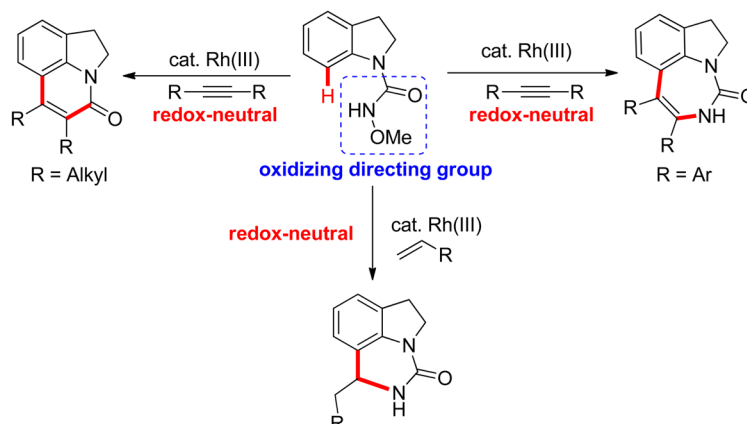
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Scheme 1. C7-Selective C–H Functionalization of Indolines

(a) Previous work: Directly access 7-substituted indolines



(b) This work: Directly access 6- or 7-membered 1,7-fused indolines



group.¹⁵ In this context, also with our continued interest in the Rh(III)-catalyzed C–H activation/annulation reaction,¹⁶ we herein report the Rh(III)-catalyzed redox-neutral C7-selective C–H functionalization of indolines with alkynes and alkenes under mild reaction conditions via utilization of an oxidizing-directing group strategy (Scheme 1b). More significantly, this method can directly access various valuable six- or seven-membered 1,7-fused indolines.^{2,17}

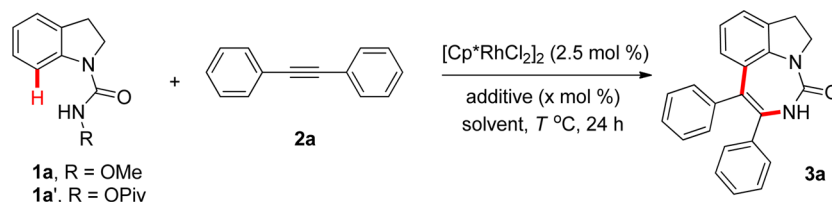
RESULTS AND DISCUSSION

We commenced our study with the coupling of *N*-methoxycarbonyl-protected indoline (**1a**) and diphenylacetylene (**2a**) (Table 1). When [Cp*₂RhCl₂]₂ (2.5 mol %)¹⁸ was employed as catalyst together with CsOAc (50 mol %) as additive at 60 °C in MeCN, the desired annulation product **3a** was obtained in 15% yield (entry 1). Changing the OMe group (**1a**) to an *O*-pivaloyl group (**1a'**), did not give any desired product **3a** (entry 2). Various solvents were screened, and MeOH was proven to be optimal, affording **3a** in 35% yield (entry 4). A stoichiometric amount (200 mol %) of CsOAc gave an improved yield of **3a** (entry 5). Moreover, the reaction efficiency could be increased at higher temperature (80 °C), affording **3a** in 65% yield (entry 6).

Finally, changing CsOAc to NaOAc gave a further improvement of yield (75%) (entry 7). In addition, a palladium catalyst system was found to be ineffective (entry 8).

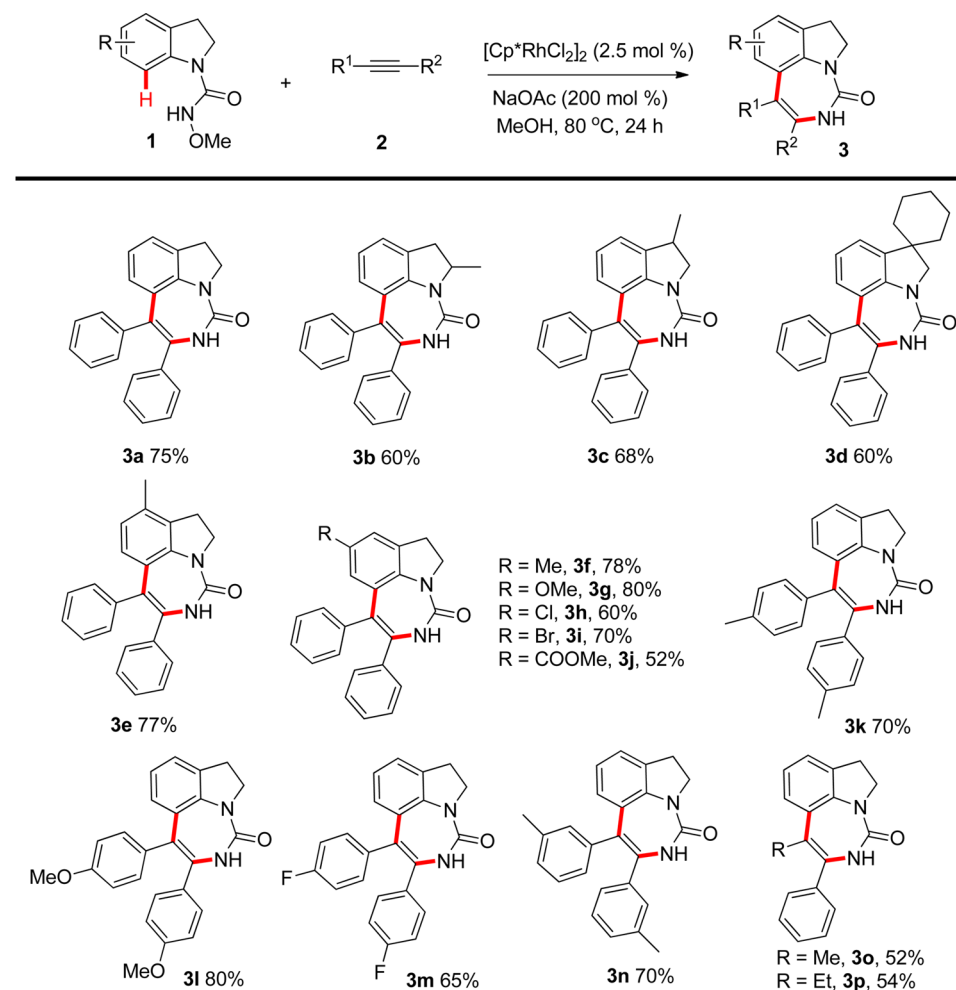
With the optimized reaction conditions in hand, the substrate scope of indolines was investigated. As shown in Scheme 2, indolines containing both electron-donating (**3b–g**) and -withdrawing (**3h–j**) groups could proceed smoothly in this transformation to give the corresponding 7-membered 1,7-fused indolines in good to high yields. Moreover, substitutions at the C2 (**3b**), C3 (**3c,d**), C4 (**3e**), and C5 (**3f–j**) were all well tolerated. Of special importance, indolines bearing chloro (**3h**), bromo (**3i**) and ester (**3j**) functional groups were also compatible with this catalytic system, thus offering the opportunity for further transformations. The scope of the alkynes partners was also explored and diphenylacetylenes bearing both electron-donating and -withdrawing groups, coupled smoothly in good to high yields (**3k–n**). Notably, the use of alkyl aryl disubstituted alkynes gave a high regioselectivity, affording **3o** and **3p** as the single regioisomer.

Next, the dialkyl-substituted alkynes were explored under the standard reaction conditions. Surprisingly, in addition to the 7-membered product **5a'**, 1,2-dihydro-4*H*-pyrrolo[3,2,1-*ij*]-quinolin-4-one **5a** was also isolated as the main product,

Table 1. Optimization of the Synthesis of Seven-Membered 1,7-Fused indolines^a

entry	substrate	additive (x mol %)	solvent	T (°C)	yield ^b (%)
1	1a	CsOAc (50 mol %)	CH ₃ CN	60	15
2	1a'	CsOAc (50 mol %)	CH ₃ CN	60	0
3	1a	CsOAc (50 mol %)	DMF	60	5
4	1a	CsOAc (50 mol %)	MeOH	60	35
5	1a	CsOAc (200 mol %)	MeOH	60	55
6	1a	CsOAc (200 mol %)	MeOH	80	65
7	1a	NaOAc (200 mol %)	MeOH	80	75
8 ^c	1a	CsOAc (200 mol %)	MeOH	80	<3

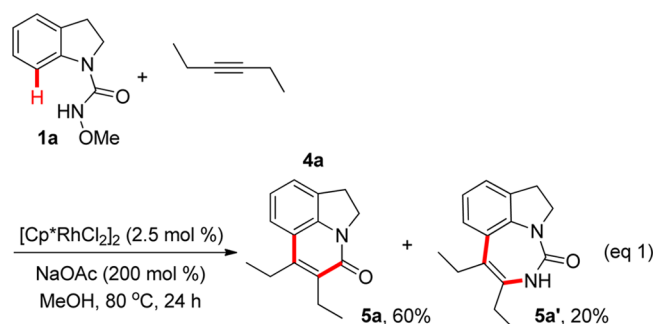
^aReaction conditions: 1a (0.2 mmol), 2a (0.4 mmol), [Cp*RhCl₂]₂ (2.5 mol %), additive (x mol %) and solvent (2 mL) in a sealed tube at 80 °C for 24 h. ^bYield of isolated product. ^cPd(OAc)₂ was used as the catalyst.

Scheme 2. Rh(III)-Catalyzed Synthesis of Seven-Membered 1,7-Fused Indolines^a

^aReactions conditions: 1 (0.20 mmol, 1.0 equiv), 2 (0.4 mmol, 2.0 equiv), [Cp*RhCl₂]₂ (2.5 mol %) and NaOAc (200 mol %) in MeOH (2.0 mL) for 24 h at 80 °C; isolated yields.

presumably due to an intramolecular nucleophilic addition of an alkenyl-Cp*Rh^{III} intermediate to carbonyl group (eq 1).

Notably, to the best of our knowledge, the intramolecular addition of alkenyl-Cp*Rh^{III} species to the less electrophilic carbonyl group is challenging and particularly rare.¹⁹ Notably,



this pyrrolo[3,2,1-*ij*]quinolin-4-one framework is known to be a CYP11B1 inhibitor, a ligand for ORL-1 receptors, and to show fungicidal activity.^{2h,20} This result encouraged us to further survey the scope of this addition reaction (Scheme 3).²¹

To our delight, a variety of electron-rich and -deficient indolines could be readily converted into 1,2,5,6-tetrahydropyrrolo[3,2,1-*ij*]quinoline-4-ones (5a–k). Importantly, the reaction also showed good compatibility with a wide range of valuable functional groups such as methoxy (5g), fluoro (5h), chloro (5i), bromo (5j), and ester (5k) groups. Tolerance to the chloro, bromo, and ester functional groups was especially noteworthy since they have been frequently used as key intermediates for further synthetic transformations. Moreover, the position of the substituent on indole moiety showed no obvious influence on the reaction outcome. For example, substitutions at the C2 (5b), C3 (5c,d), C4 (5e), C5 (5f–k) positions were all well tolerated with the current catalytic system. Interestingly, the C6-methoxy-substituted indoline gave the 7-membered product 5l' as the sole product.²² In addition, other dialkyl-substituted alkynes

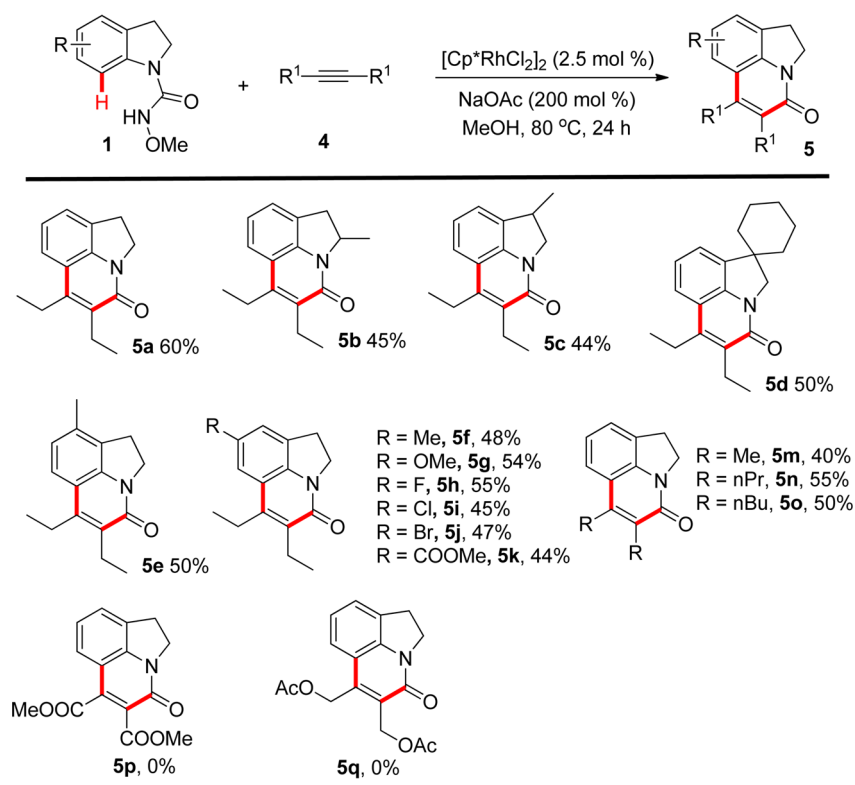
could also proceed smoothly in this transformation to give the corresponding products 5m–o. Propargyl acetates and protected propargyl alcohols did not convert into the desired products (5p,q).

Since alkenes are also ideal coupling partners,^{15c,m} the coupling of *N*-methoxycarbonyl-protected indoline (1a) and acrylate (6a) was also investigated (Table 2). When $[\text{Cp}^*\text{RhCl}_2]_2$ (2.5 mol %) was employed as catalyst together with CsOAc (50 mol %) as additive at 50 °C in MeOH, the six-membered annulation product 7a was obtained in 18% yield (entry 1). Interestingly, no seven-membered annulation product 7a' was observed. Changing of OMe (1a) to an *O*-pivaloyl group (1a') did not give any desired product 7a or 7a' (entry 2). Various solvents were tested, and DMF was proven to be optimal, affording 7a in 78% yield (entry 6).

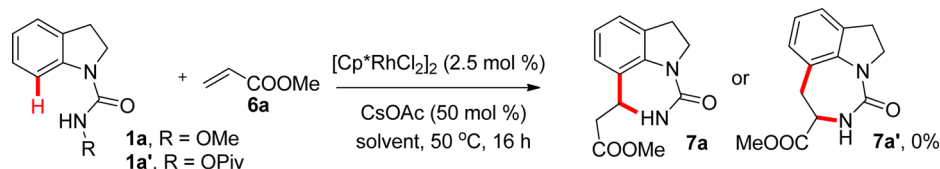
With the optimized reaction conditions in hand, the substrate scope of indolines was investigated (Scheme 4). As shown in Scheme 4, indolines containing both electron-donating (7b–d,h) and -withdrawing (7e–g) groups could proceed smoothly in this transformation to give the corresponding 1,7-fused indolines. Moreover, substitutions at the C4 (7b), C5 (7c–g), and even C6 positions (7h) were all well tolerated. Of special importance, indolines bearing chloro and bromo functional groups (7e,f), were also compatible with this catalytic system, thus offering the opportunity for further transformations.

The scope of the reaction with respect to the alkenes was also explored (Scheme 5). Satisfyingly, various acrylates such as ethyl (7i), butyl (7j), *tert*-butyl (7k), and benzyl acrylates (7l) smoothly coupled with 1a to provide the corresponding 1,7-fused indolines in good yields. In addition to acrylates, phenyl

Scheme 3. Rh(III)-Catalyzed Synthesis of 1,2-Dihydro-4H-pyrrolo[3,2,1-*ij*]quinolin-4-ones^a

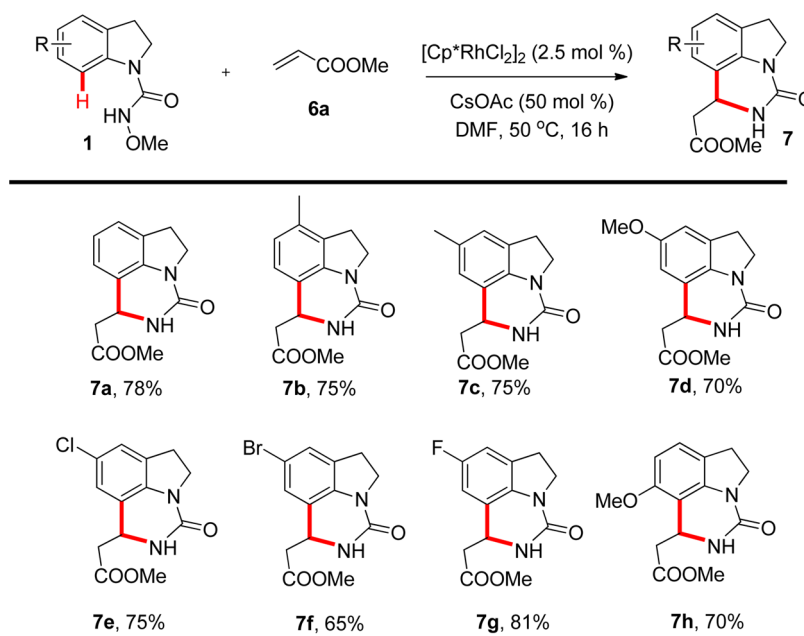


^aReaction conditions: 1 (0.20 mmol, 1.0 equiv), 4 (0.4 mmol, 2.0 equiv), $[\text{Cp}^*\text{RhCl}_2]_2$ (2.5 mol %), and NaOAc (200 mol %) in MeOH (2.0 mL) for 24 h at 80 °C; isolated yields.

Table 2. Optimization of the Rh(III)-Catalyzed Annulation Reaction with Alkenes^a

entry	substrate	solvent	yield ^b (%)
1	1a	MeOH	18
2	1a'	MeOH	0
3	1a	THF	30
4	1a	1,4-dioxane	20
5	1a	CH ₃ CN	45
6	1a	DMF	78

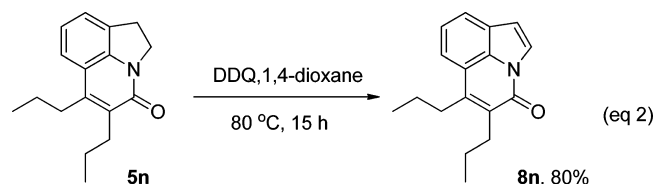
^aReaction conditions: **1a** (0.2 mmol), **6a** (0.4 mmol), $[\text{Cp}^*\text{RhCl}_2]_2$ (2.5 mol %), CsOAc (50 mol %), and solvent (2 mL) at 50 °C for 16 h. ^bYield of isolated product **7a**.

Scheme 4. Rh(III)-Catalyzed Redox-Neutral C7–H Functionalization of Various Indolines with Alkenes^a

^aReaction conditions: **1** (0.2 mmol), **6a** (0.4 mmol), $[\text{Cp}^*\text{RhCl}_2]_2$ (2.5 mol %), CsOAc (50 mol %) and DMF (2 mL) at 50 °C for 16 h. Yield of isolated product.

vinyl sulfone was also a good coupling partner, giving the desired product **7m** in 48% yield. Alkenes containing other functional groups, such as nitrile (**7n**) and phosphonate (**7o**), were also tolerated.

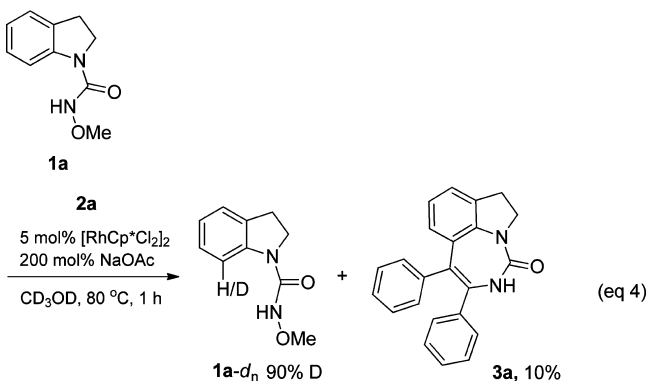
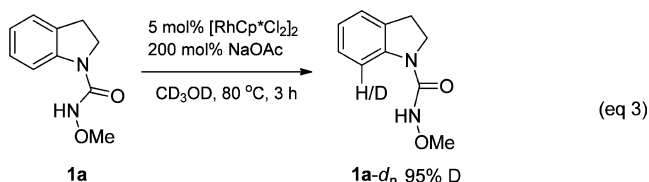
The utility of this method was further highlighted by its successful conversion into 1,7-fused indoles with DDQ. For example, the product **5n** was easily oxidized to give a pyrroloquinolinone framework of **8n** (eq 2), which is prevalent



in some biologically active indole based alkaloids²³ and whose derivatives are found to have some unusual photosensitizing properties.²⁴

For a preliminary mechanistic study, H/D exchange experiments were performed. A significant H/D scrambling was observed in the C7-position of indoline when **1a** was reacted with the $[\text{RhCp}^*\text{Cl}_2]_2$ catalyst in CD₃OD in the absence of diphenylacetylene (**2a**) (eq 3). Moreover, in the presence of **2a**, a similar deuteration was also observed in the reisolated **1a** (eq 4). Together, these results indicate the reversibility of the C–H activation.

A preliminary mechanistic pathway is postulated for the Rh(III)-catalyzed redox-neutral functionalization of aryl C–H bond with alkynes and alkenes (Scheme 6). First, a Rh(III)-catalyzed reversible C-7 C–H bond cleavage occurs to give rhodacycle **A** upon proton abstraction. Insertion of alkyne to the carbon–rhodium bond of **A** affords the eight-membered rhodacycle **B**. When aryl alkynes were involved, reductive elimination occurs with the formation of intermediates **C** and a Rh(I) species. Oxidative addition of **C** releases the product **3** and regenerates the Rh(III) catalyst. If alkyl alkynes were involved, an



intramolecular nucleophilic addition followed by release of methoxyamine, occurred to give product **5** with the regeneration of Rh(III) catalyst.

Similarly, insertion of olefin to the carbon–rhodium bond of **A** affords the eight-membered rhodacycle **E**. This metallacycle might undergo a β -hydride elimination/N–O bond cleavage to provide the intermediate **F** and regenerate the Rh(III) catalyst. Finally, an intramolecular aza-Michael addition²⁵ occurs to give 1,7-fused indoline **7**.

CONCLUSIONS

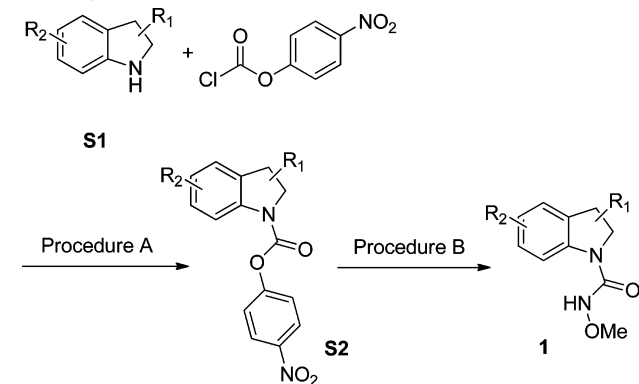
In summary, we have developed the first example of Rh(III)-catalyzed redox-neutral C7-selective C–H functionalization of indolines with alkynes and alkenes. In general, this mild method has enabled us to rapidly access valuable six- and seven-membered 1,7-fused indolines. Given the valuable structure of

the products and lack of external oxidants, this method should be of synthetic utility.

EXPERIMENTAL SECTION

General Information. ¹H NMR (400 or 300 MHz) and ¹³C NMR (125, 100 MHz) spectra were determined with CDCl₃ as solvent and tetramethylsilane (TMS) as internal standard. Chemical shifts were reported in ppm from internal TMS (δ). All coupling constants (*J* values) were reported in hertz (Hz). High-resolution mass spectra were recorded using the EI method with a double-focusing magnetic mass analyzer. Reactions were monitored by thin-layer chromatography or LC–MS analysis. Column chromatography (petroleum ether/ethyl acetate) was performed on silica gel (200–300 mesh). All of the reagents were used directly as obtained commercially unless otherwise noted.

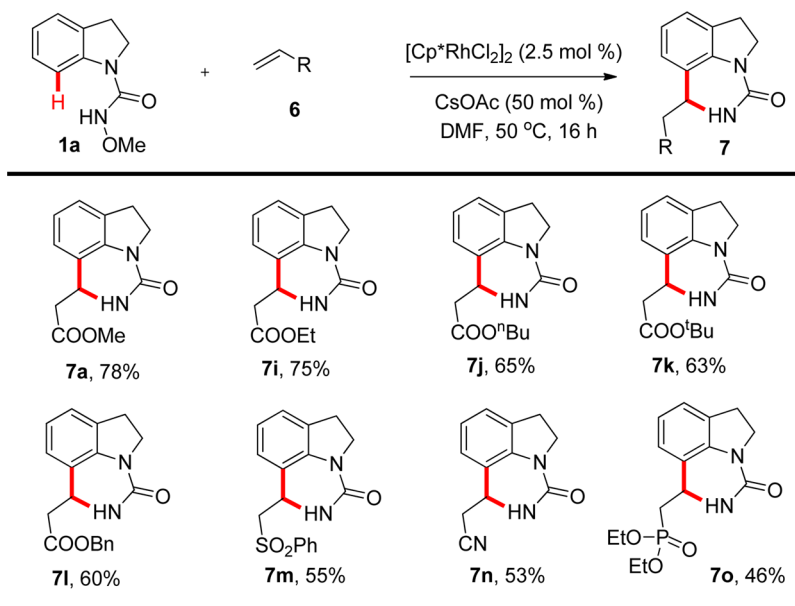
Typical Experimental Procedure for Synthesis of N-Methoxyindoline-1-carboxamide **1**.



Procedure A. To a mixture of indoline **S1** (8.50 g, 43.1 mmol) and 4-nitrophenyl chloroformate (8.69 g, 43.1 mmol) in THF (120 mL) was added pyridine (3.50 mL, 43.1 mmol) at 0 °C. The mixture was stirred at room temperature under N₂ atmosphere for 3 h. The solid was collected and washed with AcOEt to give the title compound as a pale yellow solid (14.6 g, 94%), which was used for the next step without further purification.

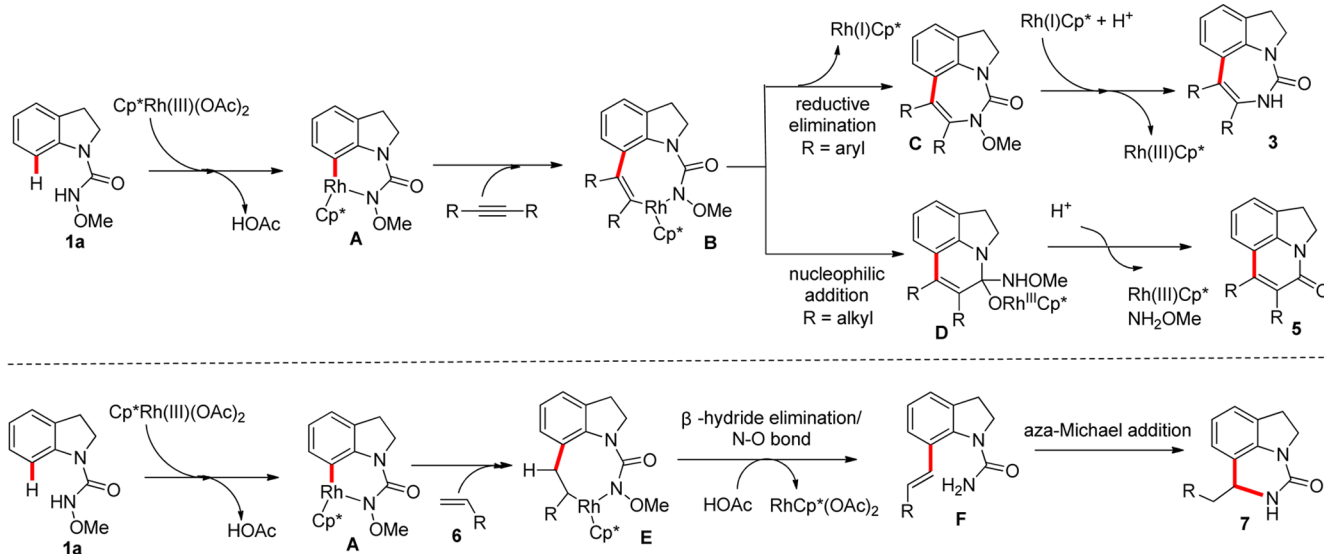
Procedure B. To a solution of compound **S2** (2.84g, 10 mmol) and methoxyamine hydrochloride (4.18 g, 50 mmol) in pyridine (50 mL)

Scheme 5. Rh(III)-Catalyzed Redox-Neutral C7–H Functionalization of Indolines with Various Alkenes^a



^aReaction conditions: **1a** (0.2 mmol), **2** (0.4 mmol), [Cp^{*}RhCl₂]₂ (2.5 mol %), CsOAc (50 mol %) and DMF (2 mL) at 50 °C for 16 h. Yield of isolated product.

Scheme 6. Proposed Mechanism for the Rh(III)-Catalyzed Indoline C7–H Functionalization with Alkynes and Alkenes



was added DBU (7.5 mL, 50 mmol) at 0 °C. The reaction mixture was stirred overnight at 60 °C. The reaction mixture was cooled, diluted with water, and extracted with AcOEt. The organic layer was washed with brine and dried over Na₂SO₄. After removal of the solvent, the residue was purified by silica gel column chromatography to give the title compound **1** as a colorless solid.

Compound 1a: column chromatography (CH₂Cl₂/CH₃OH = 30:1), 1.1 g, yield 60%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.91 (d, *J* = 8.0 Hz, 1H), 7.69 (s, 1H), 7.18 (dd, *J* = 13.4, 7.2 Hz, 2H), 6.97 (t, *J* = 7.4 Hz, 1H), 3.88 (q, *J* = 8.1 Hz, 2H), 3.81 (s, 3H), 3.17 (t, *J* = 8.5 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 155.9, 142.4, 129.9, 127.1, 124.1, 122.1, 114.6, 63.9, 45.8, 27.6; HRMS (EI) calcd for C₁₀H₁₂N₂O₂ [M]⁺ 192.0899, found 192.0895.

Compound 1b: column chromatography (CH₂Cl₂/CH₃OH = 30:1), 1.1 g, yield 60%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.70 (d, *J* = 8.0 Hz, 1H), 7.54 (s, 1H), 7.25–7.17 (m, 2H), 7.00 (t, *J* = 7.5 Hz, 1H), 4.46 (dd, *J* = 13.4, 6.7 Hz, 1H), 3.85 (s, 3H), 3.40 (dd, *J* = 15.8, 9.2 Hz, 1H), 2.65 (d, *J* = 16.0 Hz, 1H), 1.33 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 156.0, 141.2, 130.1, 127.7, 125.4, 123.0, 115.4, 64.6, 55.1, 36.4, 20.8; HRMS (EI) calcd for C₁₁H₁₄N₂O₂ [M]⁺ 206.1055, found 206.1051.

Compound 1c: column chromatography (CH₂Cl₂/CH₃OH = 30:1), 1.2 g, yield 60%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.89 (d, *J* = 8.1 Hz, 1H), 7.53 (s, 1H), 7.25–7.14 (m, 2H), 7.01 (td, *J* = 7.4, 1.0 Hz, 1H), 4.07 (t, *J* = 9.2 Hz, 1H), 3.82 (s, 3H), 3.55–3.46 (m, 1H), 3.43 (dd, *J* = 9.2, 6.7 Hz, 1H), 1.36 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 156.2, 142.2, 135.6, 127.9, 123.5, 122.9, 115.2, 64.6, 54.5, 34.9, 20.3; HRMS (EI) calcd for C₁₁H₁₄N₂O₂ [M]⁺ 206.1055, found 206.1048.

Compound 1d: column chromatography (CH₂Cl₂/CH₃OH = 30:1), 1.0 g, yield 50%, amorphous solid; ¹H NMR (300 MHz, CDCl₃) δ 7.86 (d, *J* = 8.1 Hz, 1H), 7.52 (s, 1H), 7.23–7.09 (m, 2H), 6.99 (td, *J* = 7.4, 0.9 Hz, 1H), 3.82 (s, 3H), 3.68 (s, 2H), 1.85–1.51 (m, 7H), 1.35 (t, *J* = 10.1 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 156.3, 141.6, 139.7, 127.9, 122.9, 122.5, 115.1, 64.7, 56.8, 44.8, 37.3, 25.3, 23.0; HRMS (EI) calcd for C₁₅H₂₀N₂O₂ [M]⁺ 260.1525, found 260.1547.

Compound 1e: column chromatography (CH₂Cl₂/CH₃OH = 30:1), 1.5 g, yield 55%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.72 (d, *J* = 8.0 Hz, 1H), 7.45 (s, 1H), 7.13 (t, *J* = 7.8 Hz, 1H), 6.82 (d, *J* = 7.6 Hz, 1H), 3.92 (t, *J* = 8.5 Hz, 2H), 3.83 (s, 3H), 3.09 (t, *J* = 8.5 Hz, 2H), 2.24 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 156.3, 142.4, 134.2, 129.2, 127.8, 123.8, 112.6, 64.6, 46.4, 26.9, 18.6; HRMS (EI) calcd for C₁₁H₁₄N₂O₂ [M]⁺ 206.1055, found 206.1053.

Compound 1f: column chromatography (CH₂Cl₂/CH₃OH = 30:1), 0.55 g, yield 55%, amorphous solid; ¹H NMR (300 MHz, CDCl₃) δ 7.73 (d, *J* = 8.7 Hz, 1H), 7.35 (s, 1H), 6.98 (d, *J* = 6.4 Hz, 2H), 3.86 (dd,

J = 16.8, 8.1 Hz, 2H), 3.80 (s, 3H), 3.13 (t, *J* = 8.4 Hz, 2H), 2.29 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 156.2, 140.3, 132.4, 130.5, 128.1, 125.4, 114.8, 64.6, 46.6, 28.1, 20.9; HRMS (EI) calcd for C₁₁H₁₄N₂O₂ [M]⁺ 206.1055, found 206.1046.

Compound 1g: column chromatography (CH₂Cl₂/CH₃OH = 30:1), 0.6 g, yield 58%, amorphous solid; ¹H NMR (300 MHz, CDCl₃) δ 7.78 (d, *J* = 8.5 Hz, 1H), 7.30 (s, 1H), 6.77–6.66 (m, 2H), 3.93–3.84 (m, 2H), 3.81–3.79 (m, 3H), 3.77 (s, 3H), 3.15 (t, *J* = 8.5 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 156.1, 155.8, 136.4, 131.9, 115.7, 112.1, 111.2, 64.6, 55.7, 46.6, 28.3; HRMS (EI) calcd for C₁₁H₁₄N₂O₃ [M]⁺ 222.1004, found 222.1006.

Compound 1h: column chromatography (CH₂Cl₂/CH₃OH = 30:1), 0.7 g, yield 52%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.88 (dd, *J* = 9.6, 4.7 Hz, 1H), 7.38 (s, 1H), 6.95–6.85 (m, 2H), 3.97–3.89 (m, 2H), 3.82 (s, 3H), 3.20 (t, *J* = 8.6 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 158.9 (d, *J* = 275.1), 156.1, 138.95, 132.0 (d, *J* = 19.8), 115.9 (d, *J* = 19.5), 113.8 (d, *J* = 54.6), 111.9 (d, *J* = 57.9), 64.6, 46.7, 28.1; HRMS (EI) calcd for C₁₀H₁₁FN₂O₂ [M]⁺ 210.0805, found 210.0801.

Compound 1i: column chromatography (CH₂Cl₂/CH₃OH = 30:1), 0.6 g, yield 58%, amorphous solid; ¹H NMR (300 MHz, CDCl₃) δ 7.84 (d, *J* = 8.3 Hz, 1H), 7.38 (s, 1H), 7.14 (d, *J* = 8.8 Hz, 2H), 3.89 (t, *J* = 8.6 Hz, 2H), 3.80 (s, 3H), 3.16 (t, *J* = 8.5 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 156.1, 141.6, 132.1, 127.7, 127.6, 124.8, 116.2, 64.7, 46.6, 27.9; HRMS (EI) calcd for C₁₀H₁₁ClN₂O₂ [M]⁺ 226.0509, found 226.0501.

Compound 1j: column chromatography (CH₂Cl₂/CH₃OH = 30:1), 0.7 g, yield 52%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.82 (d, *J* = 8.4 Hz, 1H), 7.45 (s, 1H), 7.32–7.29 (m, 1H), 3.90 (t, *J* = 8.6 Hz, 2H), 3.82 (s, 3H), 3.19 (t, *J* = 8.6 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 156.1, 142.1, 132.5, 130.5, 127.6, 116.6, 115.1, 64.6, 46.5, 27.9; HRMS (EI) calcd for C₁₀H₁₁BrN₂O₂ [M]⁺ 270.0004, found 270.0001.

Compound 1k: column chromatography (CH₂Cl₂/CH₃OH = 30:1), 0.7 g, yield 52%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 8.02–7.91 (m, 2H), 7.85 (s, 1H), 7.46 (d, *J* = 10.1 Hz, 1H), 4.00–3.93 (m, 2H), 3.91 (s, 3H), 3.84 (d, *J* = 0.5 Hz, 3H), 3.25 (t, *J* = 8.6 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 166.9, 156.1, 147.1, 130.4, 130.3, 126.1, 125.9, 124.4, 115.8, 114.5, 64.6, 51.9, 46.8, 27.6; HRMS (EI) calcd for C₁₂H₁₄N₂O₄ [M]⁺ 250.0954, found 250.0951.

Compound 1l: column chromatography (CH₂Cl₂/CH₃OH = 30:1), 0.4 g, yield 50%, amorphous solid; ¹H NMR (300 MHz, CDCl₃) δ 7.59 (d, *J* = 2.1 Hz, 1H), 7.42 (s, 1H), 7.02 (d, *J* = 8.2 Hz, 1H), 6.51 (dd, *J* = 8.3, 2.3 Hz, 1H), 3.88 (t, *J* = 8.5 Hz, 2H), 3.80 (d, *J* = 2.7 Hz, 6H), 3.10 (t, *J* = 8.5 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 159.6, 156.2, 143.9, 124.8, 122.1, 108.8, 101.5, 64.7, 55.6, 47.2, 27.4; HRMS (EI) calcd for C₁₁H₁₄N₂O₃ [M]⁺ 222.1004, found 222.1006.

General Procedure for the Synthesis of 3 (Taking 3a as an Example). (RhCp*Cl₂)₂ (2.5 mol %), NaOAc (200 mol %), indoline **1a**

(0.2 mmol), diphenylacetylene **2a** (0.4 mmol), and MeOH (2 mL, 0.1 M) were added to a sealed test tube under air. The reaction mixture was stirred at 80 °C for 24 h. The crude mixture was filtered through Celite and concentrated under reduced pressure. The residue was purified by flash column chromatography (PE:EA = 10:1) to give the desired product **3a**.

Compound 3a: 50 mg, yield 75%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.20–7.10 (m, 8H), 7.10–7.00 (m, 3H), 6.81 (t, J = 7.6 Hz, 1H), 6.54 (d, J = 7.9 Hz, 1H), 5.84 (s, 1H), 4.28 (t, J = 8.4 Hz, 2H), 3.07 (t, J = 8.4 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 162.5, 141.6, 139.7, 139.6, 136.5, 132.9, 131.7, 128.6, 128.4, 127.9, 127.7, 126.8, 124.5, 123.9, 123.6, 47.6, 27.6; HRMS (EI) calcd for C₂₃H₁₈N₂O [M]⁺ 338.1419, found 338.1417.

Compound 3b: column chromatography (PE:EA = 10:1), 42 mg, yield 60%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.23–7.07 (m, 8H), 7.08–6.98 (m, 3H), 6.82 (dd, J = 13.8, 6.2 Hz, 1H), 6.54 (d, J = 7.9 Hz, 1H), 5.82 (s, 1H), 5.11–4.93 (m, 1H), 3.40 (dd, J = 16.0, 9.5 Hz, 1H), 2.54 (dd, J = 16.1, 2.6 Hz, 1H), 1.39 (d, J = 6.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 162.3, 140.5, 139.7, 139.7, 136.5, 131.8, 131.7, 128.6, 128.4, 128.1, 127.9, 126.8, 124.8, 123.8, 123.4, 55.1, 35.2, 22.1; HRMS (EI) calcd for C₂₄H₂₀N₂O [M]⁺ 352.1576, found 352.1577.

Compound 3c: column chromatography (PE:EA = 10:1), 47 mg, yield 68%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.19–7.10 (m, 8H), 7.08–6.99 (m, 3H), 6.85 (t, J = 7.6 Hz, 1H), 6.56 (d, J = 7.9 Hz, 1H), 5.88 (s, 1H), 4.49 (dd, J = 11.4, 9.0 Hz, 1H), 3.75 (dd, J = 11.5, 7.0 Hz, 1H), 3.44–3.32 (m, 1H), 1.35 (d, J = 6.9 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 162.5, 141.1, 139.7, 139.5, 138.2, 136.5, 131.6, 128.6, 128.4, 127.9, 127.9, 127.6, 126.8, 124.0, 123.6, 123.3, 55.4, 34.4, 19.7; HRMS (EI) calcd for C₂₄H₂₀N₂O [M]⁺ 352.1576, found 352.1572.

Compound 3d: column chromatography (PE:EA = 8:1), 48 mg, yield 60%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.23–7.07 (m, 8H), 7.06–6.98 (m, 3H), 6.88–6.82 (m, 1H), 6.54 (dd, J = 7.9, 1.2 Hz, 1H), 5.84 (s, 1H), 4.10 (s, 2H), 1.74 (dd, J = 18.7, 12.4 Hz, 5H), 1.64–1.46 (m, 5H); ¹³C NMR (125 MHz, CDCl₃) δ 162.6, 142.4, 140.6, 139.8, 139.6, 136.6, 131.7, 128.6, 128.5, 127.9, 127.7, 126.8, 124.0, 123.6, 122.3, 57.6, 44.0, 36.8, 25.7, 23.2. HRMS (EI) calcd for C₂₈H₂₆N₂O [M]⁺ 406.2045, found 406.2041.

Compound 3e: column chromatography (PE:EA = 10:1), 54 mg, yield 77%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.23–7.08 (m, 8H), 7.03 (dd, J = 7.5, 2.0 Hz, 2H), 6.65 (d, J = 8.0 Hz, 1H), 6.46 (d, J = 8.0 Hz, 1H), 5.87 (s, 1H), 4.29 (t, J = 8.4 Hz, 2H), 2.97 (t, J = 8.4 Hz, 2H), 2.21 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 162.4, 141.3, 139.6, 135.5, 134.2, 131.5, 128.5, 128.3, 127.9, 127.8, 127.8, 126.6, 124.9, 124.8, 123.5, 47.3, 26.4, 18.6; HRMS (EI) calcd for C₂₄H₂₀N₂O [M]⁺ 352.1576, found 352.1574.

Compound 3f: column chromatography (PE:EA = 10:1), 54 mg, yield 78%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.23–7.09 (m, 8H), 7.09–6.99 (m, 2H), 6.90 (s, 1H), 6.36 (s, 1H), 5.88 (s, 1H), 4.26 (t, J = 8.3 Hz, 2H), 3.02 (t, J = 8.3 Hz, 2H), 2.12 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 162.6, 139.8, 139.5, 139.4, 136.5, 133.5, 133.1, 131.6, 128.6, 128.3, 128.1, 127.9, 127.8, 127.3, 126.7, 125.3, 123.6, 47.7, 27.6, 20.9; HRMS (EI) calcd for C₂₄H₂₀N₂O [M]⁺ 352.1576, found 352.1575.

Compound 3g: column chromatography (PE:EA = 6:1), 58 mg, yield 80%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.19–7.04 (m, 8H), 7.06–7.00 (m, 2H), 6.71–6.61 (m, 1H), 6.09 (d, J = 2.5 Hz, 1H), 5.84 (s, 1H), 4.31–4.22 (m, 2H), 3.58 (s, 3H), 3.03 (t, J = 8.3 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 162.4, 156.5, 139.7, 139.3, 136.9, 135.2, 134.3, 131.6, 128.6, 128.4, 128.3, 127.9, 126.8, 123.3, 113.1, 110.4, 55.7, 47.9, 28.1; HRMS (EI) calcd for C₂₄H₂₀N₂O [M]⁺ 368.1525, found 368.1517.

Compound 3h: column chromatography (PE:EA = 10:1), 44 mg, yield 60%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.12 (dd, J = 9.2, 5.7 Hz, 8H), 7.01 (dd, J = 6.6, 4.1 Hz, 3H), 6.51 (s, 1H), 5.88 (s, 1H), 4.28 (t, J = 8.4 Hz, 2H), 3.05 (t, J = 8.4 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 162.1, 140.2, 139.5, 138.8, 137.5, 134.9, 131.6, 129.2, 128.9, 128.5, 128.2, 127.4, 127.2, 124.5, 122.6, 47.9, 27.6; HRMS (EI) calcd for C₂₃H₁₇ClN₂O [M]⁺ 372.1029, found 372.1024.

Compound 3i: column chromatography (PE:EA = 10:1), 58 mg, yield 70%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.13 (ddd,

J = 10.0, 8.5, 3.9 Hz, 9H), 7.00 (dd, J = 7.3, 2.1 Hz, 2H), 6.65 (s, 1H), 5.90 (s, 1H), 4.27 (t, J = 8.4 Hz, 2H), 3.05 (t, J = 8.4 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 162.1, 140.8, 139.4, 138.8, 137.6, 135.2, 131.5, 130.3, 129.4, 128.5, 128.2, 128.2, 127.3, 127.2, 122.5, 116.7, 47.8, 27.5; HRMS (EI) calcd for C₂₃H₁₇BrN₂O [M]⁺ 416.0524, found 416.0522.

Compound 3j: column chromatography (PE:EA = 8:1), 41 mg, yield 52%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.71 (d, J = 1.1 Hz, 1H), 7.28 (s, 1H), 7.21–7.08 (m, 8H), 7.01 (dd, J = 7.5, 1.9 Hz, 2H), 5.95 (s, 1H), 4.31 (t, J = 8.5 Hz, 2H), 3.76 (s, 3H), 3.09 (t, J = 8.5 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 166.6, 161.5, 145.7, 139.4, 138.9, 136.9, 133.3, 131.5, 130.3, 128.5, 128.5, 128.2, 128.1, 127.3, 127.1, 125.9, 125.6, 123.0, 52.1, 47.9, 27.1; HRMS (EI) calcd for C₂₅H₂₀N₂O₃ [M]⁺ 396.1474, found 396.1481.

Compound 3k: column chromatography (PE:EA = 10:1), 51 mg, yield 70%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.14–6.89 (m, 9H), 6.81 (t, J = 7.6 Hz, 1H), 6.56 (d, J = 7.9 Hz, 1H), 5.84 (s, 1H), 4.27 (t, J = 8.3 Hz, 2H), 3.06 (t, J = 8.3 Hz, 2H), 2.25 (d, J = 4.3 Hz, 6H); ¹³C NMR (125 MHz, CDCl₃) δ 162.9, 141.8, 137.6, 137.1, 136.7, 136.3, 136.3, 132.9, 131.5, 129.1, 128.7, 128.4, 128.1, 127.9, 124.3, 123.8, 123.4, 47.5, 27.7, 21.4, 21.3, 0.14; HRMS (EI) calcd for C₂₅H₂₂N₂O [M]⁺ 366.1732, found 366.1730.

Compound 3l: column chromatography (PE:EA = 7:1), 63 mg, yield 80%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.06 (t, J = 7.2 Hz, 3H), 6.93 (d, J = 8.6 Hz, 2H), 6.81 (t, J = 7.6 Hz, 1H), 6.72–6.65 (m, 4H), 6.57 (d, J = 7.8 Hz, 1H), 5.83 (s, 1H), 4.26 (t, J = 8.4 Hz, 2H), 3.74 (d, J = 2.3 Hz, 6H), 3.05 (t, J = 8.3 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 162.9, 158.9, 158.2, 141.7, 136.1, 132.8, 132.7, 132.5, 132.1, 129.9, 128.1, 127.9, 124.3, 123.8, 123.1, 113.7, 113.4, 55.3, 55.2, 47.5, 27.7; HRMS (EI) calcd for C₂₅H₂₂N₂O₃ [M]⁺ 398.1630, found 398.1628.

Compound 3m: column chromatography (PE:EA = 10:1), 48 mg, yield 65%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.16–7.03 (m, 3H), 6.97 (dd, J = 8.5, 5.6 Hz, 2H), 6.92–6.74 (m, 5H), 6.50 (d, J = 7.8 Hz, 1H), 5.88 (s, 1H), 4.26 (t, J = 8.4 Hz, 2H), 3.06 (t, J = 8.3 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 163.1 (d, J = 34.4 Hz), 162.2, 160.6 (d, J = 32.5 Hz), 141.5, 135.7, 135.6 (d, J = 3.5 Hz), 135.3 (d, J = 3.3 Hz), 133.2 (d, J = 8.3 Hz), 133.1, 130.5 (d, J = 8.1 Hz), 127.8, 127.3, 124.7, 123.9, 122.9, 115.7, 115.5, 115.3, 115.0, 47.6, 27.6; HRMS (EI) calcd for C₂₃H₁₆F₂N₂O [M]⁺ 374.1231, found 374.1228.

Compound 3n: column chromatography (PE:EA = 10:1), 51 mg, yield 70%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.10–6.98 (m, 4H), 6.93 (dd, J = 11.6, 7.1 Hz, 3H), 6.83 (dd, J = 17.3, 9.8 Hz, 3H), 6.58 (d, J = 7.9 Hz, 1H), 5.86 (s, 1H), 4.27 (t, J = 8.4 Hz, 2H), 3.06 (t, J = 8.4 Hz, 2H), 2.20 (d, J = 6.4 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 162.7, 141.7, 139.6, 139.5, 137.9, 137.4, 136.5, 132.9, 132.2, 129.1, 128.7, 128.6, 128.1, 128.0, 127.9, 127.7, 127.5, 125.8, 124.4, 123.8, 123.5, 47.5, 27.7, 21.4; HRMS (EI) calcd for C₂₅H₂₂N₂O [M]⁺ 366.1732, found 366.1731.

Compound 3o: column chromatography (PE:EA = 12:1), 28 mg, yield 52%, amorphous solid; ¹H NMR (400 MHz, CDCl₃) δ 7.46–7.33 (m, 5H), 7.14–7.09 (m, 2H), 7.03 (t, J = 7.5 Hz, 1H), 5.58 (s, 1H), 4.31–4.16 (m, 2H), 3.03 (t, J = 8.3 Hz, 2H), 1.98 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 163.6, 142.3, 139.6, 135.2, 132.9, 129.2, 128.8, 128.6, 125.4, 124.3, 124.2, 116.8, 47.3, 27.8, 19.0; HRMS (EI) calcd for C₁₈H₁₆N₂O [M]⁺ 276.1263, found 276.1255.

Compound 3p: column chromatography (PE:EA = 12:1), 31 mg, yield 54%, amorphous solid; ¹H NMR (300 MHz, CDCl₃) δ 7.46–7.28 (m, 5H), 7.13 (t, J = 8.0 Hz, 2H), 7.03 (t, J = 7.5 Hz, 1H), 5.51 (s, 1H), 4.24 (t, J = 8.2 Hz, 2H), 3.03 (t, J = 8.3 Hz, 2H), 2.43 (q, J = 7.4 Hz, 2H), 0.83 (t, J = 7.4 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 163.8, 143.4, 139.5, 135.2, 133.2, 128.8, 128.8, 128.4, 126.7, 125.0, 124.1, 123.5, 47.1, 27.8, 24.3, 14.1; HRMS (EI) calcd for C₁₉H₁₈N₂O [M]⁺ 290.1419, found 290.1412.

General Procedure for the Synthesis of 5 and 6 (taking 5a and 5a' as an Example). (RhCp*Cl₂)₂ (2.5 mol %), NaOAc (200 mol %), indoline **1a** (0.2 mmol), 3-hexyne **4a** (0.4 mmol), and MeOH (2 mL, 0.1 M) were added to a test tube under air. The reaction mixture was stirred at 80 °C for 24 h. The crude mixture was filtered through Celite and concentrated under reduced pressure. The residue was purified by flash column chromatography (PE:EA = 15:1) to give the compound

Sa'. Subsequently, the major product was further eluted with PE/EA (3:1) to provide the pure compound **5a**.

Compound 5a: column chromatography (PE:EA = 3:1), 27 mg, yield 60%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.45 (dd, $J = 8.1$, 0.8 Hz, 1H), 7.28–7.24 (m, 1H), 7.13 (dd, $J = 8.0$, 7.2 Hz, 1H), 4.42 (dd, $J = 8.7$, 7.6 Hz, 2H), 3.44–3.32 (m, 2H), 2.88 (q, $J = 7.6$ Hz, 2H), 2.75 (q, $J = 7.5$ Hz, 2H), 1.26 (t, $J = 7.6$ Hz, 3H), 1.17 (t, $J = 7.5$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 161.1, 146.2, 141.0, 133.9, 130.8, 123.8, 122.9, 120.9, 118.1, 47.0, 27.2, 21.7, 20.5, 14.4, 14.1; HRMS (EI) calcd for $\text{C}_{15}\text{H}_{17}\text{NO}$ $[\text{M}]^+$ 227.1310, found 227.1308.

Compound 5a': column chromatography (PE:EA = 15:1), 10 mg, yield 20%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.06 (dd, $J = 10.1$, 4.1 Hz, 2H), 6.98 (t, $J = 7.5$ Hz, 1H), 5.31 (s, 1H), 4.21–4.13 (m, 2H), 2.99 (t, $J = 8.3$ Hz, 2H), 2.45 (q, $J = 7.5$ Hz, 2H), 2.26 (q, $J = 7.5$ Hz, 2H), 1.19 (t, $J = 7.5$ Hz, 3H), 1.07 (t, $J = 7.5$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 164.7, 143.2, 136.9, 133.0, 128.1, 124.5, 124.1, 123.6, 121.6, 47.1, 27.9, 27.9, 24.3, 15.1, 12.7; HRMS (EI) calcd for $\text{C}_{15}\text{H}_{18}\text{N}_2\text{O}$ $[\text{M}]^+$ 242.1419, found 242.1417.

Compound 5b: column chromatography (PE:EA = 3:1), 21 mg, yield 45%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.48–7.44 (m, 1H), 7.24 (d, $J = 1.0$ Hz, 1H), 7.14 (dd, $J = 8.0$, 7.2 Hz, 1H), 5.10–4.98 (m, 1H), 3.59 (dd, $J = 16.6$, 9.4 Hz, 1H), 2.95 (dd, $J = 16.6$, 3.7 Hz, 1H), 2.88 (q, $J = 7.6$ Hz, 2H), 2.75 (qd, $J = 7.5$, 3.3 Hz, 2H), 1.60 (d, $J = 6.4$ Hz, 3H), 1.27 (t, $J = 7.6$ Hz, 3H), 1.17 (t, $J = 7.5$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 160.9, 145.9, 140.4, 134.5, 129.4, 123.9, 122.9, 121.0, 117.9, 56.7, 36.4, 21.7, 20.8, 20.5, 14.4, 14.1; HRMS (EI) calcd for $\text{C}_{16}\text{H}_{19}\text{NO}$ $[\text{M}]^+$ 241.1467, found 241.1465.

Compound 5c: column chromatography (PE:EA = 3:1), 21 mg, yield 44%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.47 (d, $J = 8.0$ Hz, 1H), 7.25 (d, $J = 5.7$ Hz, 1H), 7.16 (t, $J = 7.6$ Hz, 1H), 4.59 (dd, $J = 12.6$, 9.4 Hz, 1H), 3.97 (dd, $J = 12.6$, 5.5 Hz, 1H), 3.74 (dd, $J = 15.5$, 6.7 Hz, 1H), 2.89 (q, $J = 7.6$ Hz, 2H), 2.76 (q, $J = 7.4$ Hz, 2H), 1.44 (d, $J = 7.0$ Hz, 3H), 1.26 (t, $J = 7.6$ Hz, 3H), 1.18 (t, $J = 7.5$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 161.0, 146.2, 140.4, 136.0, 134.0, 123.0, 122.9, 121.2, 117.9, 54.9, 34.8, 21.8, 20.9, 20.5, 14.5, 14.1; HRMS (EI) calcd for $\text{C}_{16}\text{H}_{19}\text{NO}$ $[\text{M}]^+$ 241.1467, found 241.1464.

Compound 5d: column chromatography (PE:EA = 5:1), 29 mg, yield 50%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.47 (d, $J = 7.9$ Hz, 1H), 7.23 (d, $J = 7.1$ Hz, 1H), 7.17 (t, $J = 7.6$ Hz, 1H), 4.24 (s, 2H), 2.89 (q, $J = 7.6$ Hz, 2H), 2.76 (q, $J = 7.4$ Hz, 2H), 1.85–1.31 (m, 10H), 1.28 (d, $J = 7.6$ Hz, 3H), 1.18 (t, $J = 7.4$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 161.0, 146.2, 140.0, 139.8, 133.9, 122.9, 121.9, 121.3, 118.0, 57.8, 45.7, 37.9, 25.4, 23.1, 21.8, 20.5, 14.5, 14.1; HRMS (EI) calcd for $\text{C}_{20}\text{H}_{25}\text{NO}$ $[\text{M}]^+$ 295.1936, found 295.1934.

Compound 5e: column chromatography (PE:EA = 3:1), 24 mg, yield 50%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.37 (d, $J = 8.2$ Hz, 1H), 6.96 (d, $J = 8.2$ Hz, 1H), 4.46–4.37 (m, 2H), 3.32–3.22 (m, 2H), 2.86 (q, $J = 7.6$ Hz, 2H), 2.74 (q, $J = 7.4$ Hz, 2H), 2.34 (s, 3H), 1.24 (t, $J = 7.6$ Hz, 3H), 1.17 (t, $J = 7.4$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 161.3, 146.2, 140.9, 134.0, 132.6, 128.9, 124.6, 121.1, 115.9, 47.4, 26.3, 21.7, 20.4, 18.6, 14.5, 14.1; HRMS (EI) calcd for $\text{C}_{16}\text{H}_{19}\text{NO}$ $[\text{M}]^+$ 241.1467, found 241.1462.

Compound 5f: column chromatography (PE:EA = 3:1), 23 mg, yield 48%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.23 (s, 1H), 7.11 (s, 1H), 4.53–4.33 (m, 2H), 3.34 (t, $J = 8.0$ Hz, 2H), 2.86 (q, $J = 7.6$ Hz, 2H), 2.75 (q, $J = 7.5$ Hz, 2H), 2.44 (s, 3H), 1.25 (t, $J = 7.6$ Hz, 3H), 1.17 (t, $J = 7.5$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 160.9, 145.9, 139.2, 133.8, 132.6, 130.9, 125.3, 120.6, 117.7, 47.1, 27.2, 21.9, 21.7, 20.5, 14.5, 14.1; HRMS (EI) calcd for $\text{C}_{16}\text{H}_{19}\text{NO}$ $[\text{M}]^+$ 241.1467, found 241.1463.

Compound 5g: column chromatography (PE:EA = 2:1), 28 mg, yield 54%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.94 (s, 1H), 6.87 (s, 1H), 4.50–4.33 (m, 2H), 3.85 (s, 3H), 3.35 (t, $J = 8.0$ Hz, 2H), 2.85 (q, $J = 7.6$ Hz, 2H), 2.75 (q, $J = 7.5$ Hz, 2H), 1.25 (t, $J = 7.6$ Hz, 3H), 1.17 (t, $J = 7.4$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 160.5, 156.6, 145.5, 135.9, 134.4, 132.2, 117.9, 113.5, 103.3, 56.3, 47.2, 27.3, 21.8, 20.6, 14.2, 14.1; HRMS (EI) calcd for $\text{C}_{16}\text{H}_{19}\text{NO}_2$ $[\text{M}]^+$ 257.1416, found 257.1412.

Compound 5h: column chromatography (PE:EA = 3:1), 27 mg, yield 55%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.11 (d,

$J = 10.4$ Hz, 1H), 7.03 (d, $J = 8.2$ Hz, 1H), 4.50–4.36 (m, 2H), 3.38 (t, $J = 8.0$ Hz, 2H), 2.82 (q, $J = 7.6$ Hz, 2H), 2.74 (q, $J = 7.5$ Hz, 2H), 1.24 (t, $J = 7.6$ Hz, 3H), 1.17 (t, $J = 7.5$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 160.6 (d, $J = 15.4$ Hz), 158.4, 145.4 (d, $J = 3.4$ Hz), 137.4, 135.1, 132.5 (d, $J = 9.2$ Hz), 117.9 (d, $J = 9.1$ Hz), 112.8 (d, $J = 26.7$ Hz), 106.3 (d, $J = 24.8$ Hz), 47.3, 27.2, 21.9, 20.6, 14.2, 13.9; HRMS (EI) calcd for $\text{C}_{15}\text{H}_{16}\text{FNO}$ $[\text{M}]^+$ 245.1216, found 245.1212.

Compound 5i: column chromatography (PE:EA = 3:1), 23 mg, yield 45%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.42 (s, 1H), 7.23 (s, 1H), 4.55–4.33 (m, 2H), 3.37 (t, $J = 8.0$ Hz, 2H), 2.83 (q, $J = 7.6$ Hz, 2H), 2.74 (q, $J = 7.5$ Hz, 2H), 1.25 (t, $J = 7.6$ Hz, 3H), 1.17 (t, $J = 7.5$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 160.7, 145.2, 139.7, 135.2, 132.5, 128.3, 124.4, 120.7, 118.5, 47.2, 27.1, 21.7, 20.6, 14.4, 13.9; HRMS (EI) calcd for $\text{C}_{15}\text{H}_{16}\text{ClNO}$ $[\text{M}]^+$ 261.0920, found 261.0917.

Compound 5j: column chromatography (PE:EA = 3:1), 28 mg, yield 47%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.60–7.55 (m, 1H), 7.36 (dd, $J = 2.8$, 1.3 Hz, 1H), 4.42 (dd, $J = 8.7$, 7.5 Hz, 2H), 3.37 (t, $J = 8.1$ Hz, 2H), 2.83 (q, $J = 7.6$ Hz, 2H), 2.74 (q, $J = 7.5$ Hz, 2H), 1.24 (t, $J = 7.6$ Hz, 3H), 1.16 (t, $J = 7.5$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 160.7, 145.1, 139.9, 135.2, 132.8, 127.0, 123.7, 119.1, 115.6, 47.1, 27.1, 21.7, 20.6, 14.4, 13.9; HRMS (EI) calcd for $\text{C}_{15}\text{H}_{16}\text{BrNO}$ $[\text{M}]^+$ 305.0415, found 305.0408.

Compound 5k: column chromatography (PE:EA = 2:1), 25 mg, yield 44%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.23 (s, 1H), 7.93 (s, 1H), 4.50–4.38 (m, 2H), 3.94 (s, 3H), 3.40 (t, $J = 8.1$ Hz, 2H), 2.92 (q, $J = 7.6$ Hz, 2H), 2.75 (q, $J = 7.4$ Hz, 2H), 1.27 (t, $J = 7.6$ Hz, 3H), 1.17 (t, $J = 7.5$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 167.2, 161.2, 146.4, 144.3, 134.7, 131.1, 124.9, 124.7, 124.4, 117.1, 52.4, 47.4, 26.9, 21.7, 20.5, 14.6, 13.9; HRMS (EI) calcd for $\text{C}_{17}\text{H}_{19}\text{NO}_3$ $[\text{M}]^+$ 285.1365, found 285.1363.

Compound 5l: Column chromatography (PE:EA = 10:1), 27 mg, yield 35%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.02 (d, $J = 8.1$ Hz, 1H), 6.54 (d, $J = 8.1$ Hz, 1H), 5.28 (s, 1H), 4.20 (brs, 2H), 3.81 (s, 3H), 2.93 (t, $J = 7.8$ Hz, 2H), 2.53 (d, $J = 6.3$ Hz, 2H), 2.30 (s, 2H), 1.18 (t, $J = 7.5$ Hz, 3H), 0.92 (t, $J = 7.4$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 166.4, 156.1, 146.6, 137.8, 125.2, 123.6, 123.6, 117.4, 106.5, 55.8, 47.6, 27.4, 27.3, 23.4, 14.8, 12.7; HRMS (EI) calcd for $\text{C}_{16}\text{H}_{20}\text{N}_2\text{O}_2$ $[\text{M}]^+$ 272.1525, found 272.1521.

Compound 5m: column chromatography (PE:EA = 3:1), 16 mg, yield 40%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.42 (d, $J = 8.1$ Hz, 1H), 7.29–7.24 (m, 1H), 7.17–7.10 (m, 1H), 4.48–4.33 (m, 2H), 3.38 (t, $J = 8.1$ Hz, 2H), 2.40 (s, 3H), 2.25 (s, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 161.2, 140.7, 140.4, 130.5, 128.6, 123.8, 122.8, 120.8, 118.9, 46.9, 27.2, 14.8, 13.2; HRMS (EI) calcd for $\text{C}_{13}\text{H}_{13}\text{NO}$ $[\text{M}]^+$ 199.0997, found 199.0994.

Compound 5n: column chromatography (PE:EA = 3:1), 26 mg, yield 55%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.43 (d, $J = 7.4$ Hz, 1H), 7.31–7.22 (m, 1H), 7.13 (dd, $J = 8.0$, 7.2 Hz, 1H), 4.46–4.37 (m, 2H), 3.39 (t, $J = 8.1$ Hz, 2H), 2.86–2.79 (m, 2H), 2.77–2.64 (m, 2H), 1.69–1.51 (m, 4H), 0.99 (t, $J = 7.2$ Hz, 3H), 0.95 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 161.2, 144.9, 141.0, 133.1, 130.8, 123.8, 122.8, 121.2, 118.4, 46.9, 30.8, 29.5, 27.3, 23.4, 22.9, 14.7, 14.7; HRMS (EI) calcd for $\text{C}_{17}\text{H}_{21}\text{NO}$ $[\text{M}]^+$ 255.1623, found 255.1618.

Compound 5o: column chromatography (PE:EA = 3:1), 28 mg, yield 50%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.43 (dd, $J = 8.1$, 0.7 Hz, 1H), 7.28–7.25 (m, 1H), 7.13 (dd, $J = 8.0$, 7.2 Hz, 1H), 4.47–4.32 (m, 2H), 3.38 (t, $J = 8.1$ Hz, 2H), 2.88–2.78 (m, 2H), 2.78–2.66 (m, 2H), 1.64–1.39 (m, 8H), 0.97 (dt, $J = 12.3$, 7.2 Hz, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 161.2, 145.1, 140.9, 133.0, 130.8, 123.8, 122.9, 121.1, 118.4, 46.9, 32.2, 31.8, 28.5, 27.3, 27.2, 23.4, 23.3, 14.2, 14.1; HRMS (EI) calcd for $\text{C}_{19}\text{H}_{25}\text{NO}$ $[\text{M}]^+$ 283.1936, found 283.1931.

General Procedure for the Synthesis of 7 (Taking 7a). (RhCp^*Cl_2)₂ (2.5 mol %), CsOAc (50 mol %), indoline **1a** (0.2 mmol), methyl acrylate **6a** (0.4 mmol), and DMF (2 mL, 0.1 M) were added to a test tube under air. The reaction mixture was stirred at 50 °C for 16 h. The crude mixture was filtered through Celite and concentrated under reduced pressure. The residue was purified by flash column chromatography to give the desired product **7a**. The structure was established via NOE studies (see the Supporting Information).

Compound 7a: column chromatography (PE:acetone = 2:1), 38 mg, yield 78%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.08 (dd, $J = 7.9, 5.7$ Hz, 1H), 6.92–6.83 (m, 2H), 5.59 (s, 1H), 5.06 (ddd, $J = 8.8, 4.6, 2.5$ Hz, 1H), 4.04–3.95 (m, 2H), 3.72 (s, 3H), 3.17 (t, $J = 8.6$ Hz, 2H), 2.85–2.67 (m, 2H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 171.5, 153.4, 140.6, 128.7, 124.5, 122.8, 122.8, 116.8, 52.1, 51.3, 45.8, 42.9, 28.0; HRMS (EI) calcd for $\text{C}_{13}\text{H}_{14}\text{N}_2\text{O}_3$ [M] $^+$ 246.1004, found 246.1001.

Compound 7b: Column chromatography (PE:acetone = 2:1), 39 mg, yield 76% (dr = 2.5:1), amorphous solid. The indicated compounds were separated from the reaction crude mixture by flash column chromatography on silica gel. The two compounds are having the same R_f values, and they were obtained as an inseparable mixture with a ratio of 2.5:1. The following $^1\text{H NMR}$ data are not complete due to overlapping of some of the peaks. Major product: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.06–6.84 (m, Ar–H, 3H), 5.47 (s, 1H, N–H), 5.12 (d, $J = 9.9$ Hz, 1H), 4.55 (m, 1H), 3.74 (s, 3H), 3.38 (m, 1H), 2.77–2.69 (m, 3H), 1.44 (d, $J = 6.3$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 171.7, 152.7, 139.9, 127.5, 124.7, 122.7, 122.5, 116.6, 55.8, 52.2, 51.0, 42.3, 36.9, 20.9. Minor product: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.06–6.84 (m, 3H), 5.43 (s, 1H, N–H), 5.01 (dt, $J = 10.0, 3.2$ Hz, 1H), 4.55 (m, 1H), 3.72 (s, 3H), 3.38 (m, 1H), 2.91–2.85 (m, 3H), 1.51 (d, $J = 6.3$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 171.5, 153.6, 140.4, 127.6, 124.4, 122.9, 122.8, 116.9, 55.3, 52.1, 51.3, 43.1, 37.1, 22.0; HRMS (EI) calcd for $\text{C}_{14}\text{H}_{16}\text{N}_2\text{O}_3$ [M] $^+$ 260.1161, found 260.1162.

Compound 7c: Column chromatography (PE:acetone = 2:1), 37 mg, yield 72% (dr = 1:1), amorphous solid. The indicated compounds were separated from the reaction crude mixture by flash column chromatography on silica gel. The two compounds are having the same R_f values and they were obtained as an inseparable mixture with a ratio of 1:1: $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.07–6.87 (m, Ar–H, 3H), 5.53 (s, 1H, N–H), 5.07 (m, 1H), 4.17 (m, 1H), 3.73 (s, 3H), 3.53 (m, 2H), 2.75 (m, 2H), 1.36 (d, $J = 6.4$ Hz, 3H); $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.07–6.87 (m, Ar–H, 3H), 5.53 (s, 1H, N–H), 5.07 (m, 1H), 4.17 (m, 1H), 3.72 (s, 3H), 3.53 (m, 2H), 2.75 (m, 2H), 1.36 (d, $J = 6.4$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 171.6 (171.5), 153.5 (153.2), 140.2 (139.9), 133.9 (133.7), 123.5 (123.4), 122.9, 116.8, 54.1 (53.8), 52.1 (52.1), 51.4 (51.3), 43.1 (42.9), 35.6 (35.5), 20.9 (19.8); HRMS (EI) calcd for $\text{C}_{14}\text{H}_{16}\text{N}_2\text{O}_3$ [M] $^+$ 260.1161, found 260.1158.

Compound 7d: column chromatography (PE:acetone = 2:1), 39 mg, yield 75%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.79 (d, $J = 7.8$ Hz, 1H), 6.71 (d, $J = 7.8$ Hz, 1H), 5.44 (s, 1H), 5.05 (t, $J = 5.6$ Hz, 1H), 4.02 (m, 2H), 3.73 (s, 3H), 3.10 (t, $J = 8.5$ Hz, 2H), 2.22 (s, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 171.6, 153.4, 140.3, 134.6, 127.5, 123.8, 123.0, 114.3, 52.2, 51.3, 45.8, 43.1, 27.1, 18.4; HRMS (EI) calcd for $\text{C}_{14}\text{H}_{16}\text{N}_2\text{O}_3$ [M] $^+$ 260.1161, found 260.1155.

Compound 7e: column chromatography (PE:acetone = 2:1), 39 mg, yield 75%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.91 (s, 1H), 6.68 (s, 1H), 5.42 (s, 1H), 5.17–4.91 (m, 1H), 4.10–3.87 (m, 2H), 3.73 (s, 3H), 3.14 (t, $J = 8.5$ Hz, 2H), 2.85–2.64 (m, 2H), 2.28 (s, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 171.6, 153.4, 138.4, 132.6, 128.8, 125.3, 123.1, 116.5, 52.2, 51.4, 45.9, 43.0, 28.1, 21.3; HRMS (EI) calcd for $\text{C}_{14}\text{H}_{16}\text{N}_2\text{O}_3$ [M] $^+$ 260.1161, found 260.1157.

Compound 7f: column chromatography (PE:acetone = 1:1), 38 mg, yield 70%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.70 (s, 1H), 6.44 (s, 1H), 5.36 (s, 1H), 5.12–4.88 (m, 1H), 4.06–3.96 (m, 2H), 3.75 (s, 3H), 3.74 (s, 3H), 3.16 (t, $J = 8.5$ Hz, 2H), 2.82–2.54 (m, 2H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 171.5, 156.6, 153.4, 134.4, 129.9, 117.1, 111.3, 108.3, 56.2, 52.2, 51.5, 46.1, 42.9, 28.3; HRMS (EI) calcd for $\text{C}_{14}\text{H}_{16}\text{N}_2\text{O}_4$ [M] $^+$ 276.1110, found 276.1108.

Compound 7g: column chromatography (PE/acetone = 2:1), 42 mg, yield 75%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.06 (s, 1H), 6.88 (s, 1H), 5.49 (s, 1H), 5.13–4.98 (m, 1H), 4.15–3.93 (m, 2H), 3.74 (s, 3H), 3.17 (t, $J = 8.6$ Hz, 2H), 2.75 (qd, $J = 17.1, 6.8$ Hz, 2H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 171.3, 153.0, 139.5, 130.5, 127.7, 125.0, 123.0, 117.8, 52.3, 51.1, 46.2, 42.8, 27.9; HRMS (EI) calcd for $\text{C}_{13}\text{H}_{13}\text{ClN}_2\text{O}_3$ [M] $^+$ 280.0615, found 280.0612.

Compound 7h: column chromatography (PE:acetone = 2:1), 42 mg, yield 65%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.21 (s, 1H), 7.02 (s, 1H), 5.48 (s, 1H), 5.12–4.88 (m, 1H), 3.99 (m, 2H), 3.74 (s, 3H), 3.18 (t, $J = 8.6$ Hz, 2H), 2.75 (m, 2H); $^{13}\text{C NMR}$ (100 MHz,

CDCl_3) δ 171.3, 152.9, 139.9, 130.9, 127.8, 125.8, 118.3, 114.8, 52.3, 50.9, 46.1, 42.8, 27.9; HRMS (EI) calcd for $\text{C}_{13}\text{H}_{13}\text{BrN}_2\text{O}_3$ [M] $^+$ 324.0110, found 324.0112.

Compound 7i: column chromatography (PE:acetone = 2:1), 42 mg, yield 81%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.82 (d, $J = 8.0$ Hz, 1H), 6.61 (d, $J = 8.9$ Hz, 1H), 5.46 (s, 1H), 5.02 (d, $J = 9.5$ Hz, 1H), 4.04 (t, $J = 8.6$ Hz, 2H), 3.73 (s, 3H), 3.17 (t, $J = 8.5$ Hz, 2H), 2.74 (m, 2H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 171.3, 159.4 (d, $J = 238.6$ Hz), 153.2, 136.8, 130.3 (d, $J = 8.7$ Hz), 117.2 (d, $J = 8.5$ Hz), 112.3 (d, $J = 24.8$ Hz), 109.7 (d, $J = 25.5$ Hz), 52.3, 51.2, 46.2, 42.7, 28.1; HRMS (EI) calcd for $\text{C}_{13}\text{H}_{13}\text{FN}_2\text{O}_3$ [M] $^+$ 264.0910, found 264.0911.

Compound 7j: column chromatography (PE:acetone = 1:1), 38 mg, yield 70%, amorphous solid; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.01 (d, $J = 8.1$ Hz, 1H), 6.38 (d, $J = 8.2$ Hz, 1H), 5.47 (s, 1H), 5.11 (d, $J = 10.7$ Hz, 1H), 4.09–3.94 (m, 2H), 3.79 (s, 3H), 3.73 (s, 3H), 3.11 (t, $J = 8.4$ Hz, 2H), 2.96–2.83 (m, 1H), 2.64 (dd, $J = 17.3, 10.7$ Hz, 1H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 172.2, 154.9, 153.3, 142.0, 124.9, 120.8, 105.1, 104.2, 55.8, 52.1, 48.6, 46.6, 41.2, 27.3; HRMS (EI) calcd for $\text{C}_{14}\text{H}_{16}\text{N}_2\text{O}_4$ [M] $^+$ 276.1110, found 276.1106.

Compound 7k: column chromatography (PE:acetone = 2:1), 39 mg, yield 75%, amorphous solid; $^1\text{H NMR}$ (300 MHz, CDCl_3) δ 7.12–7.04 (m, 1H), 6.93–6.84 (m, 2H), 5.47 (s, 1H), 5.12–5.00 (m, 1H), 4.19 (q, $J = 7.1$ Hz, 2H), 4.01 (t, $J = 8.8$ Hz, 2H), 3.18 (t, $J = 8.6$ Hz, 2H), 2.85–2.66 (m, 2H), 1.27 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 171.1, 153.4, 140.6, 128.8, 124.5, 122.8, 116.9, 61.2, 51.4, 45.9, 43.2, 28.1, 14.3; HRMS (EI) calcd for $\text{C}_{14}\text{H}_{16}\text{N}_2\text{O}_3$ [M] $^+$ 260.1161, found 260.1158.

Compound 7l: column chromatography (PE:acetone = 2:1), 37 mg, yield 65%, amorphous solid; $^1\text{H NMR}$ (300 MHz, CDCl_3) δ 7.08 (m, 1H), 6.90 (m, 2H), 5.44 (s, 1H), 5.07 (m, 1H), 4.13 (t, $J = 6.7$ Hz, 2H), 4.02 (t, $J = 8.4$ Hz, 2H), 3.19 (t, $J = 8.6$ Hz, 2H), 2.85–2.67 (m, 2H), 1.61 (m, 2H), 1.36 (m, 2H), 0.93 (t, $J = 7.3$ Hz, 3H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 171.3, 153.4, 140.7, 128.8, 124.5, 122.8, 116.9, 65.2, 51.4, 45.9, 43.2, 30.7, 28.1, 19.2, 13.8; HRMS (EI) calcd for $\text{C}_{16}\text{H}_{20}\text{N}_2\text{O}_3$ [M] $^+$ 288.1474, found 288.1471.

Compound 7m: column chromatography (PE:acetone = 2:1), 36 mg, yield 63%; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.16–7.00 (m, 1H), 6.91 (m, 2H), 5.48 (s, 1H), 5.14–4.98 (m, 1H), 4.11–3.98 (m, 2H), 3.20 (t, $J = 8.5$ Hz, 2H), 2.77–2.64 (m, 2H), 1.49 (s, 9H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 170.5, 153.5, 140.7, 128.7, 124.4, 122.9, 122.8, 117.1, 81.9, 51.5, 45.8, 44.2, 28.3, 28.1; HRMS (EI) calcd for $\text{C}_{16}\text{H}_{20}\text{N}_2\text{O}_3$ [M] $^+$ 288.1474, found 288.1469.

Compound 7n: column chromatography (PE:acetone = 2:1), 38 mg, yield 60%; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.44–7.33 (m, 5H), 7.11 (m, 1H), 6.95–6.83 (m, 2H), 5.57 (s, 1H), 5.24–5.14 (m, 2H), 5.11 (m, 1H), 4.03 (t, $J = 8.6$ Hz, 2H), 3.20 (t, $J = 8.5$ Hz, 2H), 2.95–2.72 (m, 2H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 170.9, 153.4, 140.6, 135.4, 128.8, 128.6, 128.6, 124.5, 122.9, 122.8, 116.8, 67.0, 51.4, 45.9, 43.2, 28.1; HRMS (EI) calcd for $\text{C}_{19}\text{H}_{18}\text{N}_2\text{O}_3$ [M] $^+$ 322.1317, found 322.1313.

Compound 7o: column chromatography (PE:acetone = 2:1), 36 mg, yield 55%; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.97 (d, $J = 7.6$ Hz, 2H), 7.72 (t, $J = 7.3$ Hz, 1H), 7.63 (t, $J = 7.5$ Hz, 2H), 7.10 (d, $J = 7.4$ Hz, 1H), 6.89 (t, $J = 7.5$ Hz, 1H), 6.83 (d, $J = 7.6$ Hz, 1H), 5.83 (s, 1H), 5.34 (d, $J = 9.5$ Hz, 1H), 4.03 (t, $J = 8.5$ Hz, 2H), 3.72–3.49 (m, 1H), 3.39 (d, $J = 14.0$ Hz, 1H), 3.18 (t, $J = 8.5$ Hz, 2H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 152.5, 140.7, 139.1, 134.3, 129.7, 128.9, 127.9, 124.9, 123.0, 122.6, 114.8, 63.2, 49.8, 45.8, 27.9; HRMS (EI) calcd for $\text{C}_{17}\text{H}_{16}\text{N}_2\text{O}_3\text{S}$ [M] $^+$ 328.0882, found 328.0880.

Compound 7p: column chromatography (PE:acetone = 2:1), 22 mg, yield 53%; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.16 (d, $J = 7.2$ Hz, 1H), 7.02 (d, $J = 7.6$ Hz, 1H), 6.96 (t, $J = 7.5$ Hz, 1H), 5.69 (s, 1H), 4.98 (td, $J = 6.1, 2.9$ Hz, 1H), 4.21–3.91 (m, 2H), 3.22 (t, $J = 8.5$ Hz, 2H), 2.75 (d, $J = 6.1$ Hz, 2H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 153.0, 140.4, 129.2, 125.4, 123.4, 123.2, 116.6, 115.1, 51.8, 46.0, 28.2, 28.1; HRMS (EI) calcd for $\text{C}_{12}\text{H}_{11}\text{N}_3\text{O}$ [M] $^+$ 213.0902, found 213.0897.

Compound 7q: column chromatography (PE:EA = 4:1), 29 mg, yield 46%; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.09 (t, $J = 4.1$ Hz, 1H), 6.91 (t, $J = 6.2$ Hz, 2H), 5.64 (s, 1H), 5.03 (dd, $J = 13.7, 8.2$ Hz, 1H), 4.27–4.08 (m, 4H), 4.08–3.95 (m, 2H), 3.18 (t, $J = 8.6$ Hz, 2H), 2.23 (m, 2H), 1.37 (td, $J = 7.1, 2.0$ Hz, 6H); $^{13}\text{C NMR}$ (125 MHz, CDCl_3) δ 153.2, 140.6, 128.8,

124.6, 122.9, 122.6, 117.8 (d, $J = 17.5$ Hz), 62.3 (dd, $J = 15.6, 6.5$ Hz), 50.3 (d, $J = 5.0$ Hz), 45.9, 35.6 (d, $J = 136.4$ Hz), 28.1, 16.6 (dd, $J = 5.6, 2.7$ Hz); HRMS (EI) calcd for $C_{15}H_{21}N_2O_4P$ $[M]^+$ 324.1239, found 324.1237.

General Procedure for the Oxidation of indoline. To a solution of **5n** (0.2 mmol) in 1,4-dioxane (2 mL) was added DDQ (0.4 mmol). The mixture was stirred at 80 °C for 15 h. The crude mixture was filtered through Celite and concentrated under reduced pressure. The residue was purified by flash column (PE:EA = 10:1) to give the desired product **8n**: 40 mg, yield 80%, amorphous solid; 1H NMR (400 MHz, $CDCl_3$) δ 7.95 (d, $J = 3.6$ Hz, 1H), 7.75 (d, $J = 7.6$ Hz, 1H), 7.66 (d, $J = 7.7$ Hz, 1H), 7.41 (t, $J = 7.7$ Hz, 1H), 6.83 (d, $J = 3.6$ Hz, 1H), 2.97–2.88 (m, 2H), 2.78–2.70 (m, 2H), 1.79–1.68 (m, 2H), 1.63 (ddd, $J = 13.0, 9.0, 4.9$ Hz, 2H), 1.09 (dt, $J = 22.0, 7.4$ Hz, 6H); ^{13}C NMR (100 MHz, $CDCl_3$) δ 159.8, 147.2, 132.9, 131.4, 127.9, 123.9, 123.8, 123.8, 121.5, 118.3, 110.4, 30.9, 29.5, 24.5, 22.9, 14.8, 14.6; HRMS (EI) calcd for $C_{17}H_{19}NO$ $[M]^+$ 253.1467, found 253.1465.

■ ASSOCIATED CONTENT

■ Supporting Information

Table S1; 1H and ^{13}C NMR spectra of all synthesized compounds. The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b00684.

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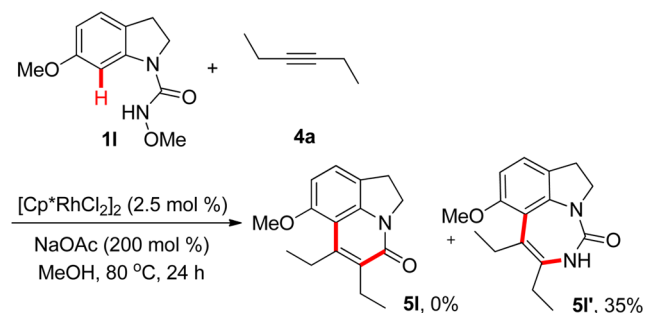
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